

Utilization of RAPD marker to analyze natural genetic variation in
Calligonum polygonoides L. - A key stone species of Thar desertSangeeta Bewal¹, Santosh Kumar Sharma², Ajay Parida³, Sadha Shivam³,
Satyawada Rama Rao^{2,*}, Arun Kumar¹¹ Cytogenetics and Molecular Biology Laboratory, Jai Narain Vyas University, Jodhpur (Raj), India² Dept. of Biotechnology and Bioinformatics, North Eastern Hill University, Shillong (Megh), India³ Plant and Molecular Biology, MS Swaminathan Research Foundation, Chennai, India

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Abstract

The species *Calligonum polygonoides* L. of Polygonaceae is one of the most economically important resources of the Indian desert, playing an important role in the lives of local populace. A great range of genetic diversity could be seen in diverse populations of this species which are spread all over western Rajasthan. DNA-based molecular markers are playing increasingly important role in the analysis of genetic diversity in wide range of plants. The present study is a first attempt to collect, compile and collate to define the existing natural genetic diversity at intra-specific and inter-populational levels in *C. polygonoides*. The present report deals with 22 genotypes of *C. polygonoides* collected from different districts of Rajasthan using random amplified polymorphic DNA (RAPD) marker. In total 100 fragments with 89 polymorphic bands amplified by 10 random primers, with the polymorphic rate of 89.30% were recorded. The dendrogram was constructed based on the matrix of Dice coefficients by unweighted pair-group mean analysis (UPGMA) using the NTSYS-PC version 1.8 exeter software. From the analysis of data it is imperative that predominantly obligate out breeding behavior of *C. polygonoides*, made these diverse accessions to spread and occupy specific geographical niches in the arid regions of Rajasthan.

Keywords: *Calligonum polygonoides*, Intra-specific variation, Genetic diversity, Genetic relatedness.**INTRODUCTION**

Calligonum polygonoides, bestowed the status of key-stone species of Indian desert, is an important source of food for sustenance during frequently occurring famines and is also valued for commercial and medicinal purposes (Bhandari, 1995). *Calligonum polygonoides* with smooth stems and branches, bearing abortive flowers and small succulent fruits, is eaten when food is scarce during famines in arid regions of India. Flowers made into bread or cooked with clarified butter or coconut oil to make a local delicacy (Bewal, 2007). Roots and thick branching stems are used as fuel (Singh *et al.*, 1996). The aqueous paste of plant acts as an antidote against the heavy doses of opium and also against poisonous effects of certain harmful plants (Singh *et al.*, 1996). *Calligonum polygonoides* form

communities with other dominating species at the top of sand dunes and stabilize the shifting sand dunes as it consists of dense network of roots.

One of the basic requirements for the sustainable and optimal utilization of the available meager resources of Indian desert region is to characterize and estimate the existing genetic diversity in natural populations. However, *C. polygonoides* did not receive the due attention it deserves as a key stone species of Indian desert. From cytological point of view there is a single report by Valovich *et al.* (1973), who proposed $2n=54$ as somatic chromosome number. No chromosome number reports for *C. polygonoides* or any other species of the genus *Calligonum* from India are available as yet. The studies on relationship and clustering of *Calligonum* species through isozyme analysis has been reported lately by Tao and Ren (2004). The quick perusal of published literature on molecular characterization indicates that the RAPD analysis to assess inter-specific relationship in *C. polygonoides* collections from China has been attempted by Ren *et al.* (2002). Realizing the importance and utility of RAPDs as ideal genetic

***Corresponding author:**

Satyawada Rama Rao, Ph.D.
Department of Biotechnology and Bioinformatics,
North Eastern Hill University,
Shillong (Meghalaya), India
Email: srrao@nehu.ac.in; srrao22@yahoo.com

1 markers, they have been used in the present
2 investigation, to assess the genetic diversity and
3 determine genetic relatedness/ distance among various
4 accessions of *C. polygonoides*, collected from different
5 regions of Rajasthan state of India.

6 MATERIALS AND METHODS

7 Twenty two accessions of *Calligonum polygonoides*
8 (Table 1 [Supplementary data]) collected from six districts
9 of Rajasthan (Jhunjhunu, Jodhpur, Jaisalmer, Bikaner,
10 Barmer and Nagaur) were analyzed for molecular
11 genetic variation among them. Fresh, young and small
12 caducous leaves from these plants were bulked so as to
13 make approximately 1-2g of leaf sample. Total
14 genomic DNA was extracted following CTAB method
15 described by Murray and Thompson (1980).

16
17 Analysis of DNA polymorphism was done in two
18 phases. The first (preliminary) phase, done on a subset
19 of three accessions, was an evaluation of 40 decamer
20 primers (Operon Technologies, Alameda, California,
21 USA). Ten primers, including Operon designated
22 primers OPA-11, OPA-14, OPA-17, OPA-18, OPA-19,
23 OPD-04, OPD-08, OPD-10, OPD-13 and OPD-17
24 showed consistent banding patterns and amplification
25 were selected for use in the second phase of study. The
26 second phase of experiments involved amplification of
27 genomic DNA with selected primers in each reaction.
28 The reaction mixture consisted of 10mM Tris HCl (pH
29 8.3), 50 mM KCl, 1.5 mM MgCl₂, 200 mM of each
30 dNTPs (Cinna. Gen. Inc.,Iran), 15 ng of each primer,
31 1.0 U *Taq* DNA polymerase (Banglore Genei, India)
32 and 40 ng genomic DNA per 25 µl reaction volume. A
33 negative control PCR tube containing all components
34 except genomic DNA was included in all the runs.
35 DNA amplification experiments were repeated three
36 times to confirm the authenticity of the results.
37 Amplification reactions were performed in a PCR
38 thermal cycler (Gene Amp PCR system 9700) - 40
39 cycles as 94°C for 1 minute at start followed by
40 denaturation at 94°C for 1 minute, annealing at 37°C
41 for 1 minute, extension at 72°C for 2 minutes and final
42 extension at 72°C for 10 minutes. The reactions ended
43 with an indefinite hold at a temperature of 4°C.

44
45 The amplification products were mixed with 2 µl of gel
46 loading dye (0.25% bromophenol blue, 25% Xylane
47 cyanol and 30% glycerol) and 10 µl of completed
48 amplification products, were electrophoresed in 1%
49 agarose gel with 0.5X TBE buffer (pH 8.0) for 3-4
50 hours supplying 50V of electricity. The 1 Kb ladder
51 (Fermentas), was taken as the standard marker in each
52 amplification reaction. The ethidium bromide stained
53 agarose gels were visualized under ultra violet light and
54 then photographed for documentation. The dendrogram
55 was constructed based on the matrix of Dice
56 coefficients by unweighted pair-group mean analysis

57 (UPGMA) using the software NTSYS-PC version 1.8
58 (Exeter software).

59 RESULTS

60 Forty oligonucleotide primers with random sequences
61 have been tested with three different concentration
62 levels of genomic DNA (20ng, 30ng and 40ng) among
63 22 accessions of *C. polygonoides*. Of these, 40ng has
64 been found to be ideal for each PCR amplification
65 reaction. Out of the forty oligonucleotide random
66 primers tested, only 10 of them have yielded
67 reproducible and discrete bands. Primers of OPA series
68 gave many unique bands in various samples while
69 primers of OPD series yielded mostly monomorphic
70 bands in various samples (Table 2 [Supplementary data]).
71 Genomic DNA was amplified in all the 22 genotypes of
72 *C. polygonoides* with these ten selected primers
73 producing a total of 100 bands representing the
74 amplification product(s), which were scored for their
75 presence (1) or absence (0). Of these 100 bands, 89
76 were polymorphic, 5 were monomorphic and 6 were
77 unique bands. This yielded an average of 8.9
78 polymorphic, 0.5 monomorphic and 0.6 unique
79 fragments per primer. Percentage of polymorphic bands
80 among amplified bands ranged from 83.3% (OPA 19,
81 OPD 10 and OPD 17) to a maximum of 100% (OPD 4,
82 OPD 8) with an average of 89.3% polymorphism
83 (Table 2). The number of amplified fragments per
84 sample ranged from 6 (OPD 17) to 17 (OPA 11) (Fig. 1
85 & Fig. 2). The size of the fragments varied from 200-
86 3050 bps in size.

87
88 A dendrogram based on UPGMA analysis grouped the
89 22 samples into three main clusters (Fig. 3 [Supplementary
90 data]). Cluster I is comprised of 10 samples collected
91 from Jaisalmer and Nagaur districts and a single
92 accession i.e. *C. polygonoides* JNVU/RI/2005/33 from
93 Barmer district. Cluster I could further be subdivided
94 into 3 groups. The first subcluster (Ia) had four samples
95 (2 from Jaisalmer and 2 from Nagaur), second
96 subcluster (Ib) consists of 3 samples (all from
97 Jaisalmer). The third subcluster (Ic) has all samples
98 from Nagaur district with an exception of
99 JNVU/RI/2005/33 which is from Barmer.

100
101 Cluster II grouped 2 samples, one from Jaisalmer and
102 the one from Bikaner and appeared to be distinct from
103 all others. The third main cluster i.e. Cluster III is
104 comprised of samples collected from Bikaner and
105 Barmer districts with one exception of
106 JNVU/RI/2005/55 which is from Jaisalmer district.
107 This cluster could further be resolved into three sub-
108 clusters. First subcluster (IIIa) consists of two samples
109 from Barmer and one from Jaisalmer. Second
110 subcluster (IIIb) contains populations collected from
111 Bikaner, while two accessions viz. JNVU/RI/2005/34

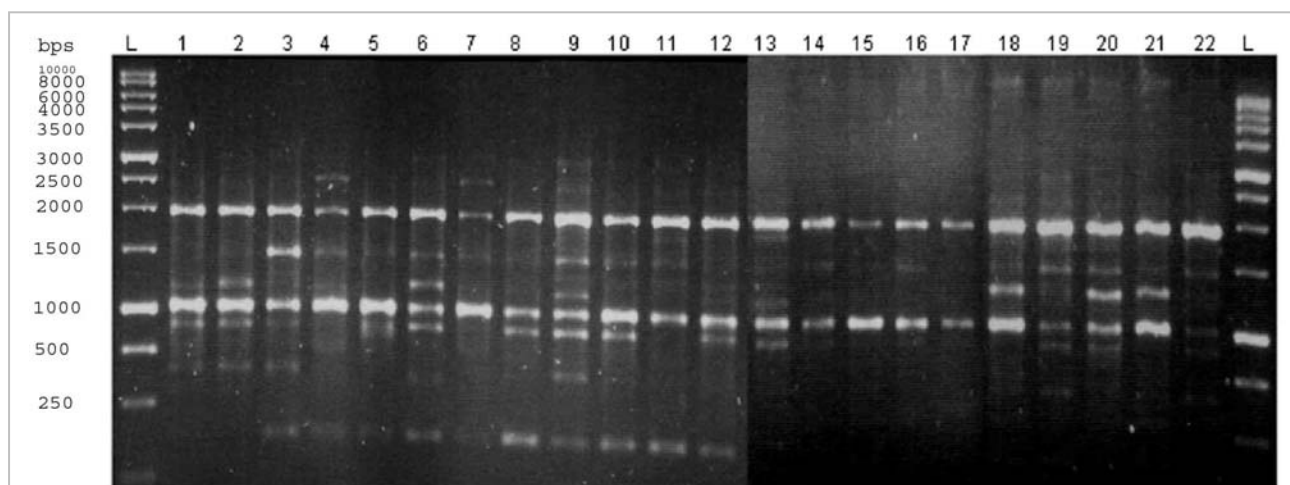


Figure 1: RAPD amplification profile of 22 accessions of *Calligonum polygonoides* using OPD 17 primer.

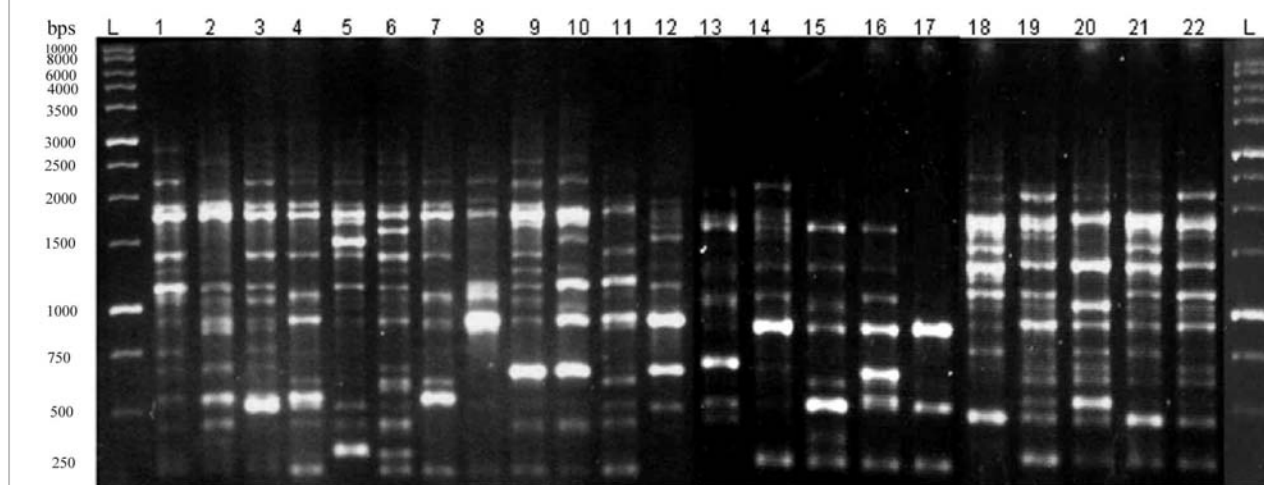


Figure 2: RAPD amplification profile of 22 accessions of *Calligonum polygonoides* using OPA 11 primer.

1 and JNVU/RI/2005/35 were grouped into third
2 subcluster (IIIc).

4 DISCUSSION

5 The minimum similarity obtained by Dice's similarity
6 coefficient was 0.51 among JNVU/RI/2005/35 and
7 JNVU/RI/2005/36 while maximum similarity was 0.93
8 among JNVU/RI/2005/36 and JNVU/RI/2005/44.

9
10 The evolution of varieties in distinct climatic zones
11 demonstrates significant levels of variations in response
12 to the selection pressure in their native environment
13 (Singh et al. 1996). It is therefore not surprising to find
14 significant levels of polymorphism among 22
15 genotypes of *C. polygonoides* as revealed by RAPD
16 markers. Clustering of genotypes was similar to some
17 extant when RAPD dendrogram was compared with the
18 morphological variation recorded in accessions
19 collected from various locations (Bewal, 2007).
20 Morphologically JNVU/RI/2005/35 collected from
21 Barmer and JNVU/RI/2005/51 collected from Jaisalmer
22 showed maximum variation with respect to plant height,

23 bark color, internodal length, flower color and fruit
24 color. The populations collected from Jaisalmer are
25 found to be most heterogenous, however on the other
26 hand populations from Bikaner seems to be
27 homogenous with very limited morphological and
28 cytological diversity being recorded in them (Bewal,
29 2007). Exceptionally JNVU/RI/2005/33 and
30 JNVU/RI/2005/37 clustered away from other samples
31 of their respective locations. This observation may be
32 attributed to marker sampling error and/or the level of
33 polymorphism detected, reinforcing again the
34 importance of the number of loci probed and their
35 coverage of the overall genome in obtaining reliable
36 estimates of genetic relationships among cultivars
37 (Loarce et al., 1996).

38
39 *Calligonum polygonoides* JNVU/RI/2005/35 was
40 collected from Pachpadra in Barmer district produced
41 many unique bands which was also reflected in
42 morphological and cytological observations of this
43 plant. This sample showed pink colored flowers and red
44 colored wings on the fruit, which gives a reddish

1 appearance rather than yellow color during flowering
 2 and fruiting. The location Pachpadra (Barmer) is
 3 known for its exceedingly rich alkalinity and soil
 4 salinity. From the present investigations it is amply
 5 clear that the genetic variation recorded in accessions
 6 collected from this particular region, may be the
 7 manifestation of stress response/tolerance, at genomic
 8 DNA level, expressed by the plants.

9
 10 Dendrogram depicted in Fig.2, did indicate the pattern
 11 of clustering according to the location from which they
 12 were collected, although there were still exceptions to it.
 13 The species under study cross pollinates (*anemophily*),
 14 as reported in many members of family Polygonaceae
 15 (Raju *et al.*, 2001) which directly reflects in variations
 16 at morphological, cytological as well as molecular
 17 levels, not withstanding the fact that the populations of
 18 the species under study are located at tens to hundreds
 19 of kilometers of distance. Accessions with the most
 20 distinct DNA profiles are likely to contain the utmost
 21 number of novel alleles (Souframanien *et al.*, 2004). It
 22 is these accessions that are likely to uncover the largest
 23 number of unique and potentially useful alleles of the
 24 genes like stress resistance/tolerance. Runo *et al.* (2004)
 25 used RAPD markers to assess genetic diversity within
 26 and between population of *Melia volkensii* in order to
 27 suggest appropriate conservation and management
 28 strategies. Study of Turkish grape germplasm by
 29 Karatas *et al.* (2008) suggests that the geographical and
 30 ecological distribution of the plants contributes to
 31 higher genetic variability. The study presented here
 32 does make an attempt to correlate the genetic variation
 33 with geographical distribution of plants with available
 34 range and quantum of natural variations in populations
 35 of *C. polygonoides*. The plants do seem to follow a
 36 trend and accessions collected from same district tend
 37 to group together. There are some accessions which
 38 deviate from this pattern which can be attributed to
 39 differential native environmental conditions.
 40 Predominantly obligate out breeding behavior of *C.*
 41 *polygonoides* beside certain non-comprehensible
 42 reasons prompted these accessions to spread and
 43 occupy specific geographical niches thereby enhancing
 44 their adaptational capabilities *vis-à-vis* native climatic
 45 conditions. The grouping may further improve if more
 46 primers are included in the study. The putatively
 47 similar bands originating for RAPDs in different
 48 individuals may not necessarily be homologous,
 49 although they may share the same size in base pairs
 50 (Liu *et al.*, 1999; Runo *et al.*, 2004). This situation may
 51 at times lead to inaccurate conclusions when
 52 calculating genetic relationships. Considering this fact,
 53 simple hybridization experiment using the polymorphic
 54 bands can confirm the extent of variation and give

55 further insight into the relationship among different
 56 accessions (Latha *et al.*, 2004; Santalla *et al.*, 1998).
 57 Nevertheless the present study is a first attempt to
 58 compile, collate, analyze and define the existing natural
 59 genetic diversity at genomic level, among the
 60 populations of *Calligonum polygonoides* collected from
 61 different locations of Rajasthan (India).

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